



Discriminating between sleep and exercise-induced fatigue using computer vision and behavioral genetics

Kelsey N. Schuch, Lakshmi Narasimhan Govindarajan, Yuliang Guo, Saba N. Baskoylu, Sarah Kim, Benjamin Kimia, Thomas Serre & Anne C. Hart


To cite this article: Kelsey N. Schuch, Lakshmi Narasimhan Govindarajan, Yuliang Guo, Saba N. Baskoylu, Sarah Kim, Benjamin Kimia, Thomas Serre & Anne C. Hart (2020) Discriminating between sleep and exercise-induced fatigue using computer vision and behavioral genetics, *Journal of Neurogenetics*, 34:3-4, 453-465, DOI: [10.1080/01677063.2020.1804565](https://doi.org/10.1080/01677063.2020.1804565)


To link to this article: <https://doi.org/10.1080/01677063.2020.1804565>

 View supplementary material [↗](#)


 Published online: 19 Aug 2020.

 Submit your article to this journal [↗](#)

 Article views: 435


 View related articles [↗](#)

 View Crossmark data [↗](#)

 Citing articles: 1 View citing articles [↗](#)



Discriminating between sleep and exercise-induced fatigue using computer vision and behavioral genetics

Kelsey N. Schuch^{a,b*}, Lakshmi Narasimhan Govindarajan^{a,c*}, Yuliang Guo^{a,c}, Saba N. Baskoylu^{a,d}, Sarah Kim^{a,d}, Benjamin Kimia^{a,e}, Thomas Serre^{a,c*} and Anne C. Hart^{a,d*} 

^aCarney Institute for Brain Science, Brown University, Providence, RI, USA; ^bDepartment of Molecular Biology, Cell Biology and Biochemistry, Brown University, Providence, RI, USA; ^cDepartment of Cognitive, Linguistic and Psychological Sciences, Brown University, Providence, RI, USA; ^dDepartment of Neuroscience, Brown University, Providence, RI, USA; ^eSchool of Engineering, Brown University, Providence, RI, USA

ABSTRACT

Following prolonged swimming, *Caenorhabditis elegans* cycle between active swimming bouts and inactive quiescent bouts. Swimming is exercise for *C. elegans* and here we suggest that inactive bouts are a recovery state akin to fatigue. It is known that cGMP-dependent kinase (PKG) activity plays a conserved role in sleep, rest, and arousal. Using *C. elegans* EGL-4 PKG, we first validate a novel learning-based computer vision approach to automatically analyze *C. elegans* locomotory behavior and an edge detection program that is able to distinguish between activity and inactivity during swimming for long periods of time. We find that *C. elegans* EGL-4 PKG function impacts timing of exercise-induced quiescent (EIQ) bout onset, fractional quiescence, bout number, and bout duration, suggesting that previously described pathways are engaged during EIQ bouts. However, EIQ bouts are likely not sleep as animals are feeding during the majority of EIQ bouts. We find that genetic perturbation of neurons required for other *C. elegans* sleep states also does not alter EIQ dynamics. Additionally, we find that EIQ onset is sensitive to age and DAF-16 FOXO function. In summary, we have validated behavioral analysis software that enables a quantitative and detailed assessment of swimming behavior, including EIQ. We found novel EIQ defects in aged animals and animals with mutations in a gene involved in stress tolerance. We anticipate that further use of this software will facilitate the analysis of genes and pathways critical for fatigue and other *C. elegans* behaviors.

ARTICLE HISTORY

Received 15 January 2020
Accepted 29 July 2020

KEYWORDS

Fatigue; *C. elegans*;
locomotion; swimming;
computer vision; quiescence



Introduction

Fatigue is a commonly experienced phenomenon that is typically defined by a feeling of exhaustion combined with decreased muscle output (Wan, Qin, Wang, Sun, & Liu, 2017). Feelings of fatigue are common after vigorous physical activity or exercise, but fatigue is also a hallmark symptom of a variety of health disorders and diseases, including cancer, mood disorders, neurodegenerative disorders, and chronic fatigue syndrome. Fatigue is not limited to vertebrates; exercise eventually drives decreased spontaneous locomotion in invertebrates as well. The molecular pathways and mechanisms involved in fatigue have not been fully delineated in any animal species.


The nematode *Caenorhabditis elegans* provides a potentially powerful model system to interrogate genetic mechanisms and cellular pathways underlying fatigue. Several methods have been developed to test the neuromuscular output of *C. elegans* during locomotion, including burrowing assays through media of varying densities (Beron *et al.*, 2015) and pillar deflection strength measuring assays (Rahman *et al.*, 2018). *Caenorhabditis elegans* are typically

grown on solid media, but swimming exercise in liquid media has been shown to be energetically costly (Laranjeiro, Harinath, Burke, Braeckman, & Driscoll, 2017). Following prolonged swimming, *C. elegans* begin to spontaneously cycle between periods of active swimming with vigorous body undulations (active bouts) and periods of immobility that lack body undulations (quiescent bouts) (Ghosh & Emmons, 2008). Because swimming is exercise for *C. elegans*, these quiescent bouts may be fatigue, as they occur after the exertion of swimming and represent a decline in muscle output. The initial vigorous swimming activity could also be a result of introduction to a new environment; however, previous studies show that *C. elegans* have a reduction in recovery crawl distance with longer swimming duration, indicating that the animals are likely fatigued (Laranjeiro *et al.*, 2017). Definition and dissection of exercise-induced quiescence (EIQ) pave the way to study conserved mechanisms fundamental to fatigue in all animals.

However, using *C. elegans* to study EIQ and fatigue requires analysis of swimming behavior over long periods of time, and therefore presents logistical and computational challenges. Several methods have been specifically developed

CONTACT Anne C. Hart  anne_hart@brown.edu  Department of Neuroscience, Brown University, 185 Meeting Street, Providence, RI, USA

*These authors contributed equally to this work.

 Supplemental data for this article can be accessed [here](#).

© 2020 Informa UK Limited, trading as Taylor & Francis Group

for automatically estimating the pose of small laboratory animals, including *C. elegans* (Gomez-Marin, Partoune, Stephens, & Louis, 2012; Jung, Aleman-Meza, Riepe, & Zhong, 2014; Patel *et al.*, 2014; Restif *et al.*, 2014). For the most part, methods developed for automatically estimating the pose of small laboratory animals rely on simple image processing (e.g. background subtraction) to extract the silhouette of a body before computing a medial axis transform. A major drawback of such methods, with non-parameterized poses, is their inability to discriminate between the front and rear ends of the body, forcing researchers to rely on simple heuristics instead (Jung *et al.*, 2014; Restif *et al.*, 2014) (e.g. by computing the direction of movement and assuming that the animal moves forward). Moreover, the performance of these methods relied heavily on certain hyperparameter choices. For example, methods which perform background subtraction are dictated by factors including the granularity of temporal sampling and thresholds for change detection. Though these are in principle automated methods, some choices for method hyperparameters can yield erroneous pose estimates, making the end user's workflow tedious. Additionally, in the context of biological research, these failures need to be detected—either automatically (Restif *et al.*, 2014) or manually (Jung *et al.*, 2014; Stephens, Johnson-Kerner, Bialek, & Ryu, 2008) to exclude the corresponding frames from further behavioral analysis. These human-in-the-loop systems suffer from two drawbacks: (i) the effective throughput of the system is conditioned on the required frequency of human intervention, and (ii) opportunistic pose tracking may lead to biases in behavioral analyses if those system failures co-occur more frequently with certain behaviors (e.g. for those behaviors that yield significant self-contact and/or self-occlusion such as omega turns and coiling). Other high-throughput computer vision systems have attempted to screen the body postures of up to 120 individual animals at a time (Swierczek, Giles, Rankin, & Kerr, 2011). However, with an increasing number of animals, the pixel resolution on each of these individuals decreases, which can yield either coarse or unreliable pose metrics.

In this study, we extended previous computer-vision work (Yang & Ramanan, 2013) with a learning-based approach [see Methods and Guo, Govindarajan, Kimia, and Serre (2018) for details] which outperforms competing solutions including a representative deep neural network that exhibits state-of-the-art accuracy for human tracking. Unlike humans, *C. elegans* lack distinctive body parts, presenting a significant challenge for deep neural networks and related approaches that rely heavily on appearance alone. Moreover, the employment of machine learning alleviates the need for manual parameter tuning, and instead yields a model that best describes an animal's posture directly from the data. This system can efficiently distinguish between periods of activity and inactivity in freely swimming *C. elegans* (Guo *et al.*, 2018)—addressing the need for high-resolution analysis that can handle both extended periods of swimming and quiescent behaviors in *C. elegans*. Unlike automated tracking systems that limit body 'pose' to just a center of

mass, our system permits fine-grained analysis of body movements and assessment of locomotion changes in *C. elegans* swimming over time and additionally handles complex postures more accurately. Here, we refer to this computer vision system as poseEIQ.

Using poseEIQ, we examined *C. elegans* locomotory behaviors and EIQ after prolonged swimming. To more efficiently determine only whether animals are active or inactive, we also developed a new behavioural analysis system called edgeEIQ, which we validated using mutant strains known to have altered EIQ (Ghosh & Emmons, 2008). With this, we describe how EIQ changes over time with extended free swimming and report previously undescribed changes in EIQ as animals age. We also determined that most EIQ bouts are not a sleep state. These new computer-vision analysis systems should be valuable for *C. elegans* researchers in any field that requires accurate assessment of locomotory behavior over extended time intervals.

Methods

Strains and maintenance

Wild type N2 Bristol, MT1072 *egl-4(n477)*, DA521 *egl-4(ad450)*, HBR227 *aptf-1(gk794)*, HBR232 *aptf-1(tm3287)*, IB16 *ceh-17(np1)*, GR1307 *daf-16(mgDf50)*, and CF1038 *daf-16(mu86)* strains were used. *C. elegans* were grown on NGM agar plates with *E. coli* OP50 bacterial food at 20°C (Brenner, 1974). Animals were obtained by selecting L4 larval stage animals; after 24 h, animals were used in assays as day 1 adult animals. For aging assays, 5-fluoro-2'-deoxyuridine (FUDR) was not used to suppress progeny production. Animals were gently serially passaged using bacteria on a pick to avoid overcrowding of plates with progeny.

Microfluidic chip preparation

PDMS microfluidic chips with 24 wells (1.6 mm wide, 0.07 mm deep, 0.4 mm gap between wells) organized in four rows and six columns were created using a custom mold. A Sylgard 184 silicone elastomer kit (Dow Chemical) was used to make PDMS, which was poured onto the mold to a thickness of 4 mm. Freshly poured PDMS was degassed in a desiccator using a vacuum until air bubbles had dissipated, then placed in a 55°C oven for 18 h to cure. PDMS was removed from the mold and cut into chips using a razor. To decrease hydrophobicity before use in swimming assays, these chips were then soaked in *E. coli* OP50 culture overnight, washed with water and ethanol, then left to dry for a week.

Kanamycin-treated *E. coli* OP50 preparation

Kanamycin-treated *E. coli* OP50 food solution was prepared as previously described (Huang, Singh, & Hart, 2017). In brief, *E. coli* OP50 was streaked onto LB agar plates and cultured overnight at 37°C. A single colony was used to inoculate 100 ml of liquid LB and cultured at 30°C shaking at 220 rpm for approximately 12 h. The culture was grown until

it reached an optical density of 2–2.5 at 550 nm and concentrated to a final OD₅₅₀ of 10 (OD determinations made using diluted cultures to stay within the linear range of the spectrophotometer). 0.2 mg/mL Kanamycin was added and the culture was placed at 4 °C for 1 week to yield a static bacterial culture. Kanamycin-treated OP50 was discarded after 6 weeks of antibiotic treatment. Immediately prior to assays, 200 microliters of kanamycin-treated OP50 was pelleted and resuspended in 300 microliters of liquid NGM.

Swimming behavioral assays

Chips were cleaned of dust and debris using laboratory tape. Microfluidic chips were then placed into 35 × 10 mm Petri dishes. Each well was loaded with kanamycin-treated OP50 in liquid NGM until a dome of the liquid droplet was visible, but no dark shadows were visible (approximately 0.5 microliters). Water was added to the Petri dish until just level with the chip surface and paraffin oil was layered over droplets and surrounding water to prevent evaporation. To conserve oil, water was used underneath paraffin oil. Before loading in individual wells, animals were picked to an unseeded plate to avoid contaminating wells with additional food. In assays with multiple genotypes, loading order always changed between trials. Animals were recorded swimming at 30 frames per second for 6 hours with a Grasshopper3 4.1MP Mono USB3 Vision camera (GS3-U3-41C6M) mounted on a Zeiss Discovery V20 microscope with a 1.25× objective providing 14.8× magnification. For image capture, we used FlyCap2 version 2.12.3.2 or SpinView version 1.13.0.33. Image resolution was 2048 × 1600 pixels, 270 × 270 pixels per chamber with approximately 1,800 pixels per animal. Representative video available at DOI: 10.5281/zenodo.3604455.

For pumping analysis, videos were recorded using the same setup but at 72.0× magnification to allow for visualization of the pharynx. For quiescent bout analysis, these videos were scaled to match the resolution of all other videos to ensure that quiescent bouts were called in a similar manner. After identification of quiescent bouts, the original videos were manually analyzed for pumping status. If the pharynx was for any reason not visible during a bout (self-occlusion or debris), those bouts were not counted (five instances).

Beat rate was manually collected by analyzing the first minute of swimming. One beat was defined as a full-body bend by the animal in both directions. The experimenter was blinded during this analysis.

Automated well detection

As a pre-processing stage, wells were automatically segmented using MATLAB's connected component algorithm (function *bwboundaries*) on the output of an edge detector (Kimia, Li, Guo, & Tamrakar, 2019). The top 24 connected components were then selected—each corresponding to a different well.

edgeIQ and activity level analysis

The behavior of each animal was analyzed for each well independently. We implemented a simple 'active' vs. 'quiescent' classifier by considering the binary output of an edge detector (Kimia *et al.*, 2019) and computing the edge difference between consecutive frames using the Jaccard index defined as: $IoU = \text{Area of Overlap} / \text{Area of Union}$, where *Area of Overlap* is the number of edge pixels present in two consecutive frames and *Area of Union* is the sum of the total number of edge pixels across consecutive frames. If the *IoU* was above a threshold $\theta = 0.9$, the behavior was set to quiescent. Otherwise, the behavior was set to active. Using edges rather than pixels yielded robustness to noise compared to simpler systems based on pixels (Restif *et al.*, 2014). A final class label was computed by voting between the two behaviors over a 1 s (30 frames) time window. The activity threshold θ was treated as a hyperparameter and a grid search strategy was employed to identify the optimal value. By systematically varying θ from 0.1 to 1 (with a step size of 0.05) and computing the Jaccard index between the estimations and manually annotated labels, we were able to automatically determine optimal θ^* that yielded the highest agreement. The annotated dataset comprised active/quiescent labels from video recordings of 8 held out animals, each video lasting 20 min at 30 frames per second. The optimal threshold value θ^* was identified to be 0.9, and yielded an agreement of 92%.

Analysis of binary quiescence data

For each hour, binary quiescence data for each animal were analyzed using MATLAB to determine fractional quiescence, time to first bout, bout number, and bout duration. First, data for each animal was iterated through to determine the start and end times for every quiescent bout across the 6-h assay. Arbitrarily, we defined the minimum duration of a quiescent bout as three seconds. Fractional quiescence for each hour was determined by summing the duration of quiescent bouts observed that hour and dividing this by time in that hour. To determine bout number per hour, the number of bouts observed in an hour was tallied. For this metric, if a bout crossed over multiple hours, this bout was counted as a fractional bout. Bout duration was determined by dividing the total time spent in quiescent bouts by the number of bouts that occurred in that hour. For bout duration, if a bout crossed over multiple hours it was counted in each hour for its full duration to more accurately portray the demographics of quiescent bouts observed during each hour. This rarely resulted in bouts being counted more than once in bout duration calculations. For fractional quiescence, if a bout crossed over multiple hours, only the portion of the bout occurring in the given hour contributed to the fractional quiescence for that hour. The time to first bout was calculated by determining the second in which the first quiescent bout for an animal occurred.

poseEIQ and kinematic analysis

The approach is based on a pose tracking algorithm developed in-house (Guo *et al.*, 2018). Briefly, nine body points were chosen along the medial axis of the animal body from head to tail. We collected 10 representative video sequences (each corresponding to a different animal) with a total length of 1200 frames (30 frames per second). Ground-truth poses were manually annotated every 10 frames by marking the location of the nine body points. To train and test our computer vision system, we used a leave-one-video out procedure. Detailed evaluation can be found in (Guo *et al.*, 2018). On average, 83% of the points were correctly detected (i.e. within a 5px radius around the ground-truth landmark). This strategy outperformed competing systems for human pose estimation, specifically the original deformable part model (Yang & Ramanan, 2013) and a leading neural network architecture for human pose estimation, the convolutional pose machine (Wei, Ramakrishna, Kanade, & Sheikh, 2016). These positional points were then used to compute metrics informative of body movements, as defined by Restif *et al.* (2014). Since these measures rely heavily on canonical reorientation of the worm with respect to its head, differentiating the head from the tail becomes particularly important. To alleviate this concern, we specifically ran a second round of tracking, using the Hungarian algorithm (Kuhn, 1955), with the initial head assignment manually checked by a trained expert. Representative video of tracking available at DOI: 10.5281/zenodo.3606717.

Statistical analysis

Statistical analysis was performed using GraphPad Prism 7.0 software (La Jolla, CA). Statistical significance in fractional quiescence, bout number, and bout duration was determined using a two-way ANOVA and Dunnett's multiple comparisons testing. Statistical significance for time to first quiescent bout was determined using Kruskal–Wallis and Dunn's multiple-comparison tests. A value of $p < 0.05$ was used to determine statistical significance (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). Error bars in figures represent the standard error of the mean.

Results

Behavioral analysis of swimming *C. elegans*

Analysis of *C. elegans* movements during swimming is critical for the analysis of diverse circuits and behavior. To further characterize *C. elegans* locomotion in liquid media, we took advantage of a new computer-vision system for analysis of swimming animals briefly described by Guo *et al.* (2018), referred to here as poseEIQ (Figure 1(A)), and assessed changes in *C. elegans* locomotory behaviors during prolonged swimming. This method addresses several problems common in current image analysis methods. With respect to background subtraction methods, the temporal granularity of sampling can lead to scenarios where the animal is construed as part of the background if it is not moving. Image

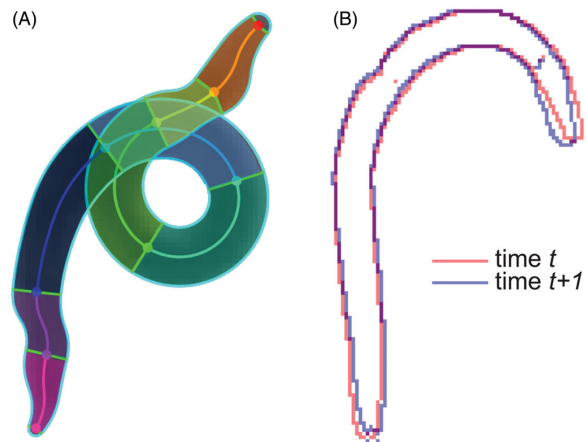


Figure 1. Visual representations of poseEIQ (A) and edgeEIQ (B). (A) poseEIQ uses a shape-consistent flexible mixture of parts model to track *C. elegans* locomotion. Image adapted from Guo *et al.* (2018). (B) edgeEIQ compares edge overlap between consecutive frames to determine activity level.

analysis tracking algorithms are also sensitive to other hyperparameter choices that can vary drastically across imaging sessions because of changes in illumination, background contrast, and imaging resolution. The poseEIQ method is a supervised machine learning algorithm, which learns appropriate configurations of body pose model parameters by optimizing an objective function that factors in both the local visual appearance of body parts and their global geometry. This alleviates the need for explicit background selection and the hand selection of hyperparameters. poseEIQ is more robust by design as long as representative image samples are included in the training phase of this model.

For this analysis, individual young adult animals were placed in small liquid drops with static bacteria as food and high-frame rate, high-resolution videos were obtained over a 6-h period. We examined both control animals and *egl-4* mutant animals, which are known to have altered locomotion. The cGMP-dependent protein kinase encoded by the *egl-4* gene is critical for a wide range of behaviors, including arousal, locomotion, and sleep (Raizen *et al.*, 2008). *egl-4(n477)* loss of function animals and *egl-4(ad450)* gain of function animals were expected to show opposing differences in locomotory behaviors, as increased EGL-4 function promotes quiescent behaviour in swimming animals (Ghosh & Emmons, 2008; McCloskey, Fouad, Churgin, & Fang-Yen, 2017).

Accurate assessment of locomotory behaviors during swimming

To determine if poseEIQ detected changes in locomotion, we analyzed recordings of 12 wild type animals and 12 animals for each of the *egl-4* mutant alleles. We used poseEIQ for two time intervals within the 6-h recording window: the first 30 min of swimming (early), when quiescent bouts are unlikely to occur were compared to 30 min of swimming (late) about 4 h (228.25 ± 4.28 min) into the videos.

Behavioral outputs are for both active and quiescent animals, but outputs can easily be changed to analysis of only active or quiescent bouts. For ease of comparison to previous work, we used locomotion parameters that were previously defined in Restif *et al.* (2014) (Supplementary Table 1). To undertake analysis for these locomotion parameters, it was necessary to assign the head and tail of each animal. Initial head and tail assignments were made computationally and verified manually. As the poseEIQ system performs shape-based pose tracking, these associations were maintained over the course of the video in most scenarios, alleviating the need for further human intervention. The resulting description of locomotion parameters is shown in Figure 2 (statistics in Table 1). Swimming beat rate was manually counted to validate poseEIQ derived wave initiation rate, a similar metric. One beat was defined as a full body movement in both directions. This yielded comparable values (Supplementary Figure 1), and values obtained running the swimming tracking program CeleST on our videos were also comparable (Restif *et al.*, 2014) (Supplementary Table 2).

As expected, the locomotory behavior of *egl-4* animals differed from wild-type animals; loss and gain of function animals are expected to have a diametrically opposed impact on behaviors. For example, at both early and late time points, average wave initiation rate and activity index were decreased in *egl-4(ad450gf)* animals and increased in *egl-4(n477lf)* animals, compared to wild-type animals (Figure 2(A,B), Table 1). We also found increased curling activity in *egl-4(ad450gf)* animals and decreased curling activity in *egl-4(n477lf)* animals compared to wild type at both early and late time points (Figure 2(C); Table 1). At the early time point, differences in stretch were observed, as curvature range was found to be increased in *egl-4(ad450gf)* animals and decreased in *egl-4(n477lf)* animals when compared to wild type (Figure 2(D); Table 1). Differences from wild type were also observed in attenuation during the early time point, with increased body wave attenuation in *egl-4(ad450gf)* animals and decreased body wave attenuation in *egl-4(n477lf)* animals (Figure 2(E); Table 1). At the later time point, *egl-4(n477lf)* animals showed increased brush stroke and *egl-4(ad450gf)* animals showed decreased brush stroke compared to wild type (Figure 2(F); Table 1). We also observed a significant increase in body wave number of *egl-4(ad450gf)* animals compared to wild type (Figure 2(G); Table 1). *egl-4(n477lf)* animals at the late time point showed increased curvature range and body wave attenuation compared to wild type (Figure 2(D,E); Table 1). The large number of diametrically opposed differences observed in *egl-4* animals suggests that poseEIQ can accurately discriminate between normal and mutant locomotion.

Prolonged swimming changes locomotory behaviors

We compared locomotion parameters across time, comparing early *versus* late time points after prolonged swimming within genotypes. Wave initiation rate and activity index decreased in wild type, *egl-4(n477lf)*, and *egl-4(ad450gf)* animals over time (Figure 2(A, B); Table 1). Body wave number

increased and brush stroke decreased in wild type and *egl-4(ad450gf)* animals between early and late time points (Figure 2(F,G); Table 1). Finally, curling activity and stretch increased in wild type animals over time (Figure 2(C,D); Table 1). The differences observed in each genotype show that not only can this approach measure the effects of prolonged exercise on locomotion using poseEIQ and these parameters, but also that changes in locomotion after prolonged swimming differ based on genotype. Overall, the changes observed at the late time point are consistent with less vigorous locomotion after 4 h of swimming.

Swimming behavioral states can be represented and analyzed using binary data or unsupervised hidden Markov models (HMM)

One drawback of detecting locomotion changes using poseEIQ is that the analysis is computationally intensive and takes a substantial amount of time to run (approximately 10 s/frame for pose estimation). Therefore, we focused on quiescent behavior during prolonged swimming and developed an edge-detection program, edgeEIQ (Figure 1(B)), that more efficiently identifies EIQ bouts in swimming animals. Knowing that diminished EGL-4 activity decreases quiescent behavior after prolonged swimming (Ghosh & Emmons, 2008), we used edgeEIQ to detect quiescent bouts on the video used above. Ethograms constructed after analysis of wild type and *egl-4* mutant animals showed clear differences in EIQ bouts for each genotype. The downside of this approach was that the activity threshold was a manually selected parameter (manual optimization of the threshold for behavior to be determined quiescent and EIQ set at a minimum duration of 3 s). As an alternative, we explored using an unsupervised Hidden Markov Model (HMM) to extract latent underlying state temporal sequences of activity. This circumvented the need for an explicit selection of EIQ minimum duration or other model parameters. Modeling was done using the open-source package *hmmlearn* in Python. The two-state HMM yielded a latent state sequence that was qualitatively similar to the ethogram constructed from the manually thresholded edgeEIQ binary data (Figure 3(A)). In a three-state HMM, the state that seems to correspond best to the inactive state from the manually thresholded binary ethogram seems to be atomic in nature, and thus cannot be further decomposed. The state corresponding best to the active state from the manually thresholded binary ethogram decomposed into two states (Supplementary Figure 2(A)). We found that HMMs with more than three states resulted in degenerate latent states, i.e. states for which more than one observation is very rare. We can identify such states from analyzing the transition matrices (Supplementary Figure 2(B,C)) and locating states for which all outgoing transition probabilities are close to zero.

To explore transitions between states predicted by HMM modeling, we computed the log-transformed transition count matrices (Supplementary Figure 2(B,C)). Prior to this, the per-frame latent state sequences were clumped into bouts of length 30 s. The equivalent latent state of the bout

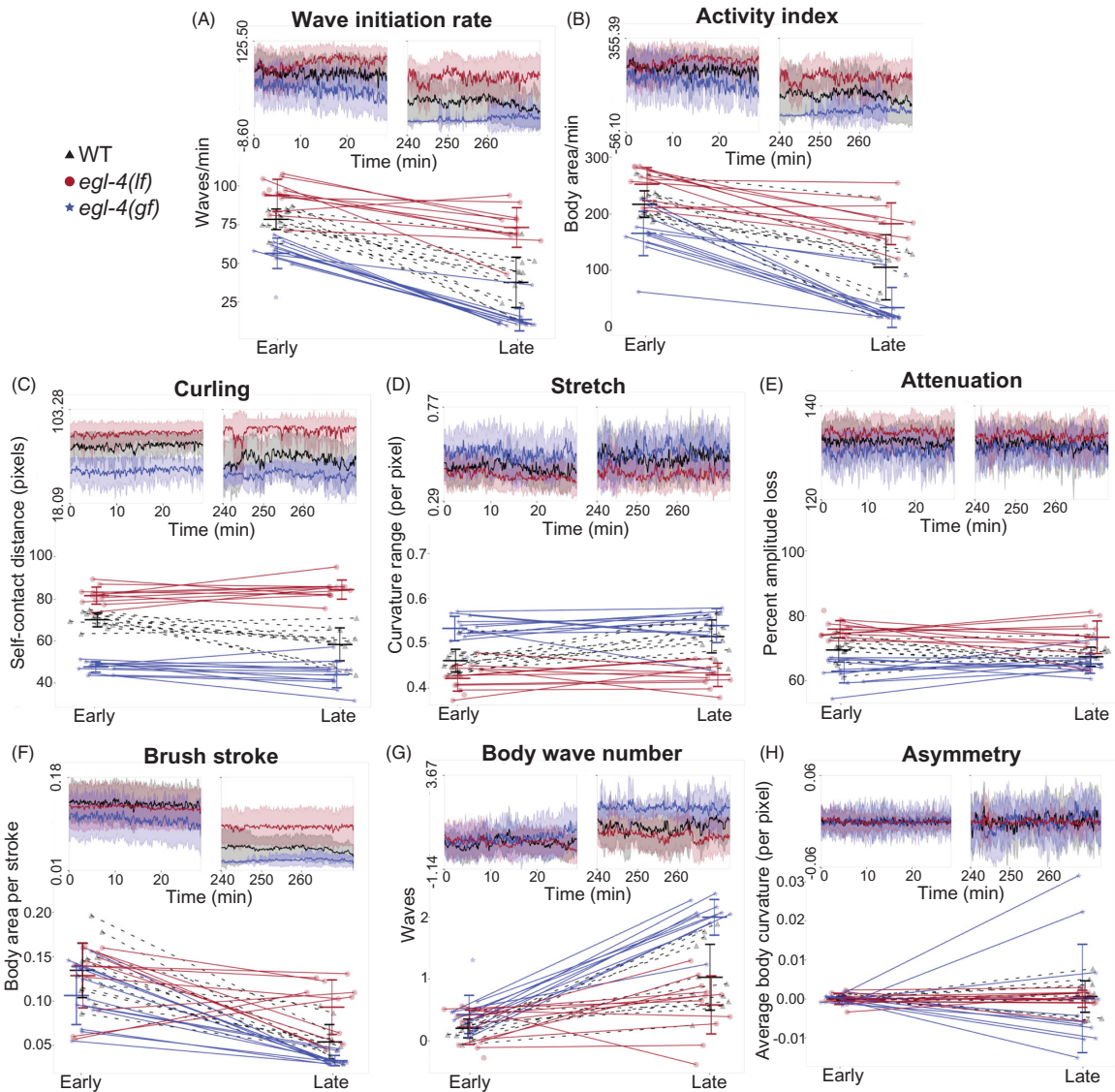


Figure 2. Locomotion analysis of *egl-4* mutant animals during prolonged swimming. Evaluation of parameters (A) Wave Initiation Rate, (B) Activity Index, (C) Curling, (D) Stretch, (E) Attenuation, (F) Brush Stroke, (G) Body Wave Number, (H) Asymmetry; each indicative of different aspects of locomotion over two windows of 30 min each. For a detailed explanation of these parameters, please refer to Restif *et al.* (2014). The ‘early’ time point is at the very beginning of the 6-h long behavioral assay while the ‘late’ time point is approximately at the 4 h mark (228.25 ± 4.28 min). The choice of these time points was motivated by the ethograms of Figure 3(A). Behavioral parameters for 12 animals from each of the three genotypes: wild type (WT), *egl-4(n477lf)* (*egl-4(lf)*), and *egl-4(ad450gf)* (*egl-4(gf)*) are shown here. The within-group mean temporal course of each parameter is shown as insets in the respective panel, with the early time point on the top left and the late time point on the top right. The average parameter value (over the 30 min window) for each individual animal is shown in the respective panel; corresponding early/late points are connected by dashed/straight lines. Activity index and brush stroke were normalized to body size (calculated in pixels). Statistical analysis available in Table 1.

Table 1. Statistical analysis of *egl-4* mutant animal locomotion parameters during prolonged swimming.

	Early		Late		Early vs. late		
	WT vs. <i>e.g.,l-4(lf)</i>	WT vs. <i>e.g.,l-4(gf)</i>	WT vs. <i>e.g.,l-4(lf)</i>	WT vs. <i>e.g.,l-4(gf)</i>	WT	<i>egl-4(lf)</i>	<i>egl-4(gf)</i>
A. Wave initiation rate	*	***	**	**	***	**	***
B. Activity index	n.s.	**	*	**	**	**	***
C. Curling	***	***	**	***	**	n.s.	n.s.
D. Stretch	*	**	*	n.s.	*	n.s.	n.s.
E. Attenuation	**	*	*	n.s.	n.s.	n.s.	n.s.
F. Brush stroke	n.s.	n.s.	*	*	***	n.s.	***
G. Body wave number	n.s.	n.s.	n.s.	**	***	n.s.	***
H. Asymmetry	n.s.	n.s.	n.s.	n.s.	n.s.	n.s.	n.s.

The non-parametric Kruskal–Wallis test (with Bonferroni correction for multiple comparisons) was used for testing significance of inter-group differences within and across time points, as well as within-group differences across time points. Error bars indicate \pm SEM. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$. WT = wild type; *egl-4(lf)* = *egl-4(n477lf)*; *egl-4(gf)* = *egl-4(ad450gf)*.

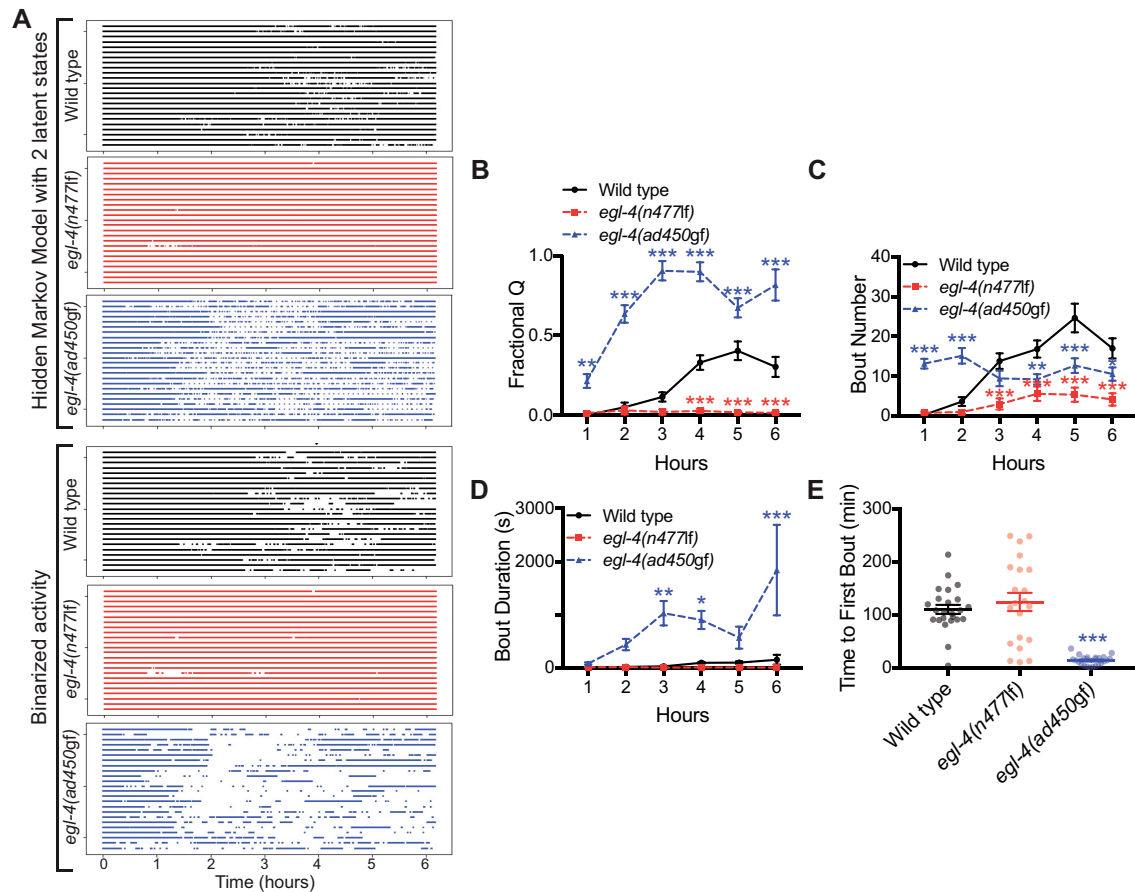


Figure 3. Analysis of exercise-induced quiescence in *egl-4* mutant animals. 24 animals per genotype. (A) Left panel: Ethograms generated using an unsupervised Hidden Markov Model for wild type, *egl-4(n477lf)*, and *egl-4(ad450gf)* animals. Each row represents the latent states of a single animal over the course of the 6 h experiment. Right panel: ethograms constructed from manually thresholded binary activity data. Filled (or empty) regions can be interpreted as an 'active' (or 'inactive') state, respectively. (B) On average, the *egl-4(n477lf)* animals showed decreased fractional quiescence (fraction of each hour spent in quiescent bouts) compared to wild type animals during hours 4 through 6. *egl-4(ad450gf)* animals showed increased average fractional quiescence at all time points compared to wild type animals. (C) *egl-4(n477lf)* animals showed decreased average number of bouts (per hour) in hours 3 through 6, compared to wild type animals. *egl-4(ad450gf)* animals showed increased average number of bouts for hours 1 and 2, and decreased average number of bouts in hours 4 through 6, compared to wild type. (D) Average duration of quiescent bouts did not differ between wild type and the *egl-4(n477lf)* animals. *egl-4(ad450gf)* animals showed increased average bout duration during hours 3, 4, and 6. Animals from three independent biological replicates (3 different days). Two-way ANOVA and Dunnett's multiple comparisons test. (E) *egl-4(ad450gf)* animals initiated quiescent bouts earlier than wild type animals, while *egl-4(n477lf)* animals were not different. Kruskal–Wallis test and Dunn's multiple comparisons test. Error bars indicate \pm SEM. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

was assigned as the statistical mode of the latent states of the constituent frames. The complete absence of state 2 in *egl-4(ad450)* gain of function animals, coupled with the relative infrequency of transitions between states 1 and 2 in the wild-type animals lends support to the aforementioned atomicity of the latent states, i.e. whether or not a given state can be decomposed further into unique latent states (Supplementary Figure 2(B,C)). The most straightforward conclusion from HMM modeling of locomotory behavior is that swimming *C. elegans* have a single inactivity state, based on activity. Additionally, swimming *C. elegans* likely have two active states, which is consistent with previous work (McCloskey *et al.*, 2017).

Using our two-state HMM, we randomly sampled body poses of animals in states that likely represent activity and quiescence (Supplementary Figure 3(A)). Similar to previous

studies, eigen decomposition revealed three modes that account for >95% of the total variance in body postures (Brown, Yemini, Grundy, Jucikas, & Schafer, 2013; Stephens *et al.*, 2008), and the statistical mean body posture of active and quiescent animals appears quite similar (Supplementary Figure 3(B,C)). Although quiescent animals sometimes assume a straight posture (Supplementary Figure 3(A)), this similarity between active and quiescent postures contrasts with previous observations reporting that quiescent animals in liquid gradually assume a rod-like posture (Ghosh & Emmons, 2008). The posture discrepancy is likely explained by differences in the definition of quiescent bouts and sampling rates. Here, short quiescent bouts are included; these were not detected or excluded in the previous study. The lack of a straight posture in a quiescent animal can be observed in the representative video available at DOI:

10.5281/zenodo.3606717. *C. elegans* display stereotypical body postures during sleep (Iwanir *et al.*, 2013; Schwarz, Spies, & Bringmann, 2012; Tramm, Oppenheimer, Nagy, Efrati, & Biron, 2014); the lack of clear difference between active and quiescent body postures here indicates that EIQ is likely distinct from sleep.

We also examined whether the duration of an active bout was related to the duration of the following quiescent bout and found that poor correlation was seen for bout durations (Spearman's $r = -0.29$) (Supplementary Figure 4(A)). This differs from the relationship between active and subsequent quiescent bouts observed during developmentally timed sleep, as a positive correlation was found between active and quiescent bout durations (Iwanir *et al.*, 2013). To explore this further, we used the raw edge-overlap data output by edgeEIQ (which is indicative of how much an animal is moving from frame to frame) to determine whether the intensity of an active bout affects the following quiescent bout. To look at animals with differing intensities of activity before entering a quiescent bout, we split animals into two groups, more active and less active, by the median activity prior to entering quiescence. We saw no difference in quiescent bout duration between the two groups (Supplementary Figure 4(B)). We also found that the less active animals returned to their initial activity level after leaving a quiescent bout—this was also true of the more active animals (Supplementary Figure 4(B)). We conclude that the relationship between active and quiescent bouts is more complicated than a linear relationship where increased activity leads to increased rest.

Accurate assessment of quiescent behavior during prolonged swimming

To confirm that our edgeEIQ program could detect previously described differences in EIQ, we compared wild-type animals swimming in the presence of food to animals carrying previously described *egl-4* mutant alleles. *egl-4(n477)* loss of function animals were predicted to show decreased EIQ and *egl-4(ad450)* gain of function animals were expected to show increased EIQ. 6-h videos of swimming animals were recorded and analyzed with the program. The fraction of time quiescent (fractional quiescence) was determined for each hour in individual animals across the 6-h experiment (Figure 3(B)). At every time point, *egl-4(ad450)* gain of function animals showed increased fractional quiescence compared to wild type, while *egl-4(n477)* loss of function animals showed decreased fractional quiescence for hours 4, 5, and 6, which is consistent with our prediction and previous work (Ghosh & Emmons, 2008). Next, we examined the average number of quiescent bouts in each hour and the average quiescence bout duration for each hour. *egl-4(n477)* animals had decreased bout numbers, starting in hour three and onward (Figure 3(C)). *egl-4(ad450)* gain of function animals showed increased bout numbers per hour during the first 2 h of swimming and decreased bout numbers for hours 4 through 6 (Figure 3(C)) and had increased bout durations at almost all time points, with the exception of hour one

(Figure 3(D)). Finally, we examined when wild type and *egl-4* mutant animals first entered a quiescent bout after prolonged swimming. On average, *egl-4(ad450)* gain of function animals showed quiescence at an earlier time than wild type animals (first quiescent bout of ≥ 3 s, Figure 3(E)). The loss of *egl-4* function did not alter quiescent bout onset. Overall, these results obtained are entirely consistent with previous work and suggest that the edgeEIQ program can robustly identify differences in EIQ and other quiescent behaviors of swimming animals.

EIQ does not require pathways necessary for developmentally timed sleep or stress-induced sleep

A well-characterized quiescent state in *C. elegans* is sleep (Hill, Mansfield, Lopez, Raizen, & Van Buskirk, 2014; Raizen *et al.*, 2008). To determine whether sleep was occurring during EIQ bouts, we manually examined quiescent bouts in wild type animals to determine if feeding was occurring. In *C. elegans*, pharyngeal pumping can be used as a metric for food intake and feeding. During each hour, pharyngeal pumping was observed in the majority of quiescent bouts; very few bouts were observed where no pharyngeal pumping occurred (Figure 4(A)). However, during several quiescent bouts, animals did not pump in the first part of the quiescent bout, then resumed pumping for the remainder of the bout. We called these 'mixed bouts' and found that the percentage of mixed bouts increased as quiescent bout duration increased (right panel, Figure 4(A)). However, in the majority of quiescent bouts animals were pumping, suggesting that most *C. elegans* EIQ bouts are not sleep.

Examination of mutant strains confirms that there is little mechanistic overlap between EIQ and previously defined *C. elegans* sleep states. Changes in EGL-4 kinase activity impact all known types of *C. elegans* sleep (Hill *et al.*, 2014; Raizen *et al.*, 2008; You, Kim, Raizen, & Avery, 2008). Perturbation of EGL-4 also alters sensory response and changes locomotion in waking animals (Figure 2). The AP2 transcription factor APTF-1 is specifically required for locomotion quiescence during *C. elegans* developmentally timed sleep (Turek, Lewandrowski, & Bringmann, 2013) and the paired homeodomain transcription factor CEH-17 is required for locomotion quiescence in stress-induced sleep (Hill *et al.*, 2014). We tested *aptf-1(gk794)* and *aptf-1(tm3287)* loss of function mutants for defects in EIQ and found that loss of APTF-1 does not alter fractional quiescence, bout number, bout duration, or time to first bout, when compared to wild type animals (Figure 4(B–E)). Although *aptf-1(tm3287)* differed from wild type in time to first bout as well as fractional quiescence and bout duration at hour five, similar changes were not observed in *aptf-1(gk794)* animals, decreasing confidence that these changes can be attributed to decreased *aptf-1* function (Figure 4(B,D,E)). We found that *ceh-17(np1)* loss of function animals were not different than wild type animals in fractional quiescence, bout number, bout duration, or time to first bout (Figure 4(F–I)). Combined, these results

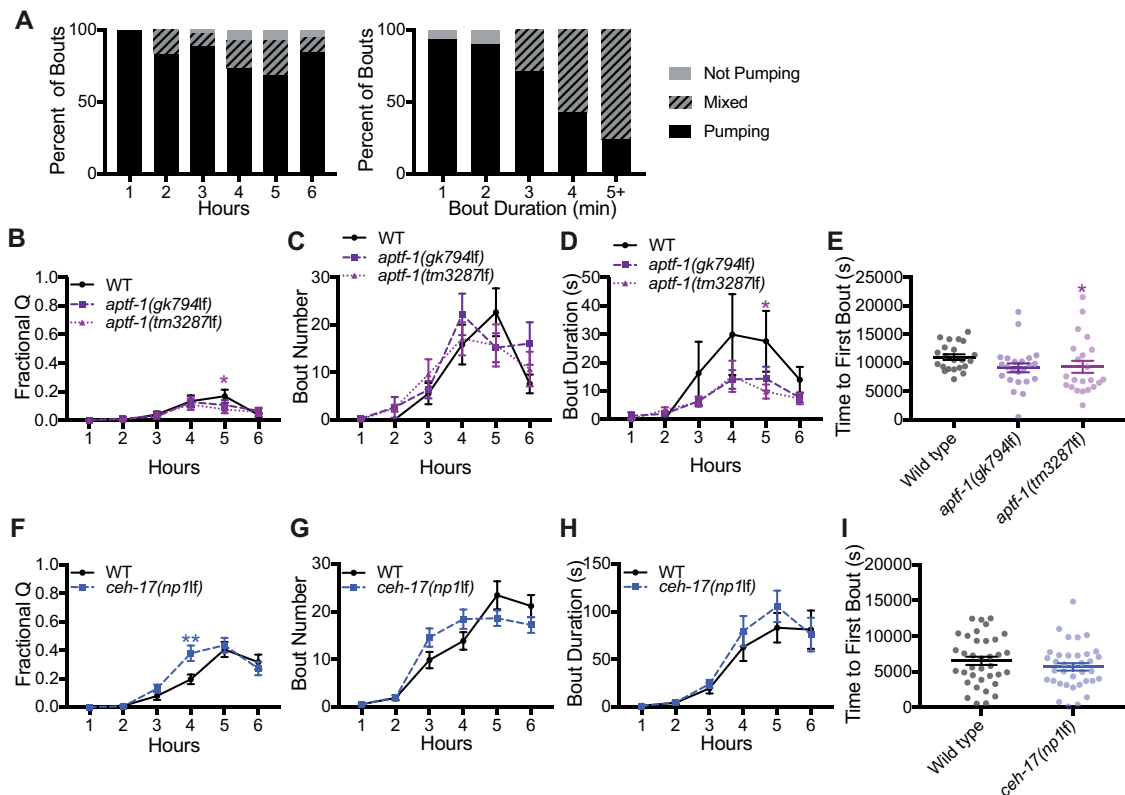


Figure 4. Exercise-induced quiescent bouts are not sleep. (A) Behavior of wild type animals during quiescent bouts was classified as ‘pumping’ (exhibited pharyngeal pumping throughout a quiescent bout), ‘mixed’ (began a quiescent bout without pumping, and resumed pumping midway through the bout), and ‘not pumping’ (no pharyngeal pumping). Left panel: The majority of quiescent bouts were classified as pumping, regardless of when they occurred. Right panel: When classified based on bout duration, most bouts lasting 3 min or less were classified as not pumping, while bouts longer than 4 min were usually classified as mixed bouts. 199 total quiescent bouts classified drawn from 5 animals. Loss of function *aptf-1(gk794lf)* and *aptf-1(tm3287lf)* animals did not differ from wild type in fractional quiescence (B), bout number (C), or bout duration (D), with the exception of fractional quiescence (B) and bout duration (D) of *aptf-1(tm3287lf)* animals at hour 5. Two-way ANOVA and Dunnett’s multiple comparisons test. (E) Average time to first bout was slightly sooner in *aptf-1(tm3287lf)*, but not *aptf-1(gk794lf)*, animals versus wild type. Kruskal-Wallis test and Dunn’s multiple comparisons test. $n = 24$ per genotype. Loss of function *ceh-17(np1lf)* showed no difference in fractional quiescence (F), bout number (G), and bout duration (H), with the exception of increased fractional quiescence at hour 5 (F). 2-way ANOVA and Dunnett’s multiple comparisons test. (I) No difference in time to first bout was observed between *ceh-17(np1lf)* animals and wild type. Kruskal-Wallis test and Dunn’s multiple comparisons test. $n = 24$ per genotype. Error bars indicate \pm SEM. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

suggest that different molecular mechanisms underlie EIQ versus *C. elegans* locomotion quiescence during sleep.

DAF-16/FOXO function delays initiation of quiescent bout cycling after prolonged swimming

The DAF-16/FOXO transcription factor plays a critical role in response to multiple stressors, including sleep restriction (Driver, Lamb, Wyner, & Raizen, 2013; Henderson & Johnson, 2001). In fasting conditions, *daf-16(mu86)* loss of function animals have total quiescent activity equivalent to wild type animals (McCloskey *et al.*, 2017). We examined both *daf-16(mgDf50)* and *daf-16(mu86)* loss of function mutant animals for changes in EIQ timing. *daf-16(mgDf50)* animals showed increased fractional quiescence at hours three, four, and six (Figure 5(A)), as well as increased bout number and bout duration at hours three and four (Figure 5(B,C)). However, these differences were not observed in

daf-16(mu86) animals. Both loss of function strains showed decreased time to first quiescent bout (Figure 5(D)). Because this last defect was seen in both mutant *daf-16* strains, we suggest that DAF-16 plays a role in the response to stress caused by prolonged swimming that is important to determining EIQ onset.

Aged animals enter quiescence at an earlier time and show increased quiescence during prolonged swimming

With age, organisms experience loss of muscle mass, known as sarcopenia, which is believed to contribute to increased frailty, decreased muscle strength, and fatigue in aged populations (Marty, Liu, Samuel, Or, & Lane, 2017). Age-related muscle deterioration has previously been observed in *C. elegans* body wall muscle (Herndon *et al.*, 2002), and aged *C. elegans* have deficits in various locomotion assays, including swim rate (Mulcahy, Holden-Dye, & O’Connor, 2013; Restif

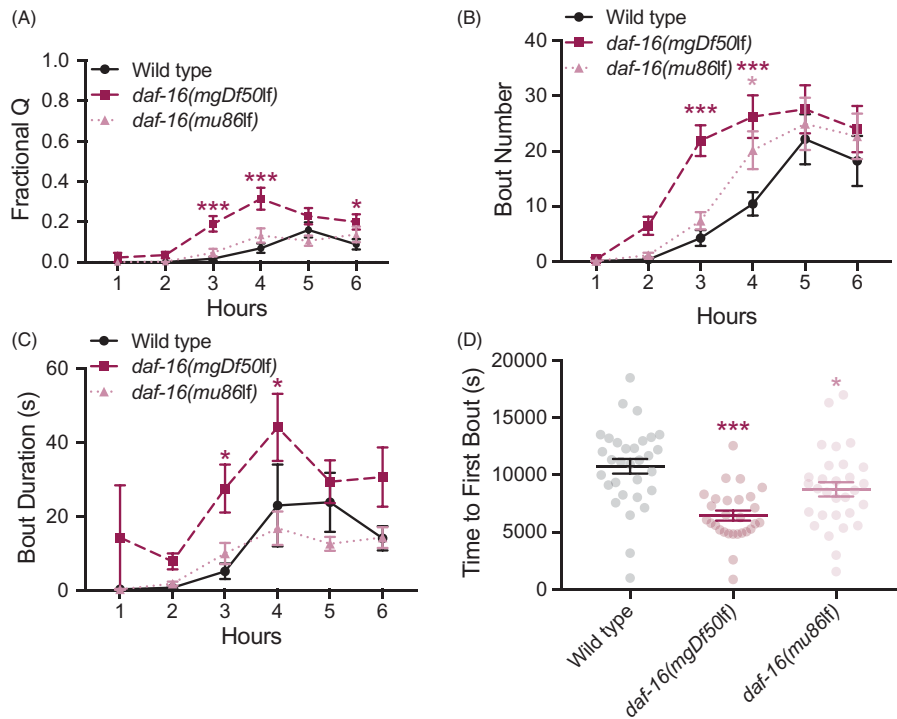


Figure 5. Analysis of exercise-induced quiescence in *daf-16* mutant animals. (A) Loss of function *daf-16(mgDf50lf)* animals showed increased average fractional quiescence, compared to wild type at hours 3, 4, and 6. This difference was not repeated in *daf-16(mu86lf)* animals. (B) *daf-16(mu86lf)* animals showed increased average number of bouts per hour, compared to wild type in hour 4, and *daf-16(mgDf50lf)* animals showed an increase during hours 3 and 4. (C) *daf-16(mgDf50lf)* animals showed increased average bout duration, compared to wild type during hours 3 and 4, while *daf-16(mu86lf)* animals showed no difference. Two-way ANOVA and Dunnett's multiple comparisons test. (D) The *daf-16(mgDf50lf)* and *daf-16(mu86lf)* animals both showed decreased average time to first bout, compared to wild type. Kruskal–Wallis test and Dunn's multiple comparisons test. $n = 32$ per genotype. Error bars indicate \pm SEM. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

et al., 2014). To test whether aged animals show defective EIQ, we aged wild type animals for 1, 2, 3, 4, 7, and 10 days into adulthood. Then, we quantified differences in EIQ timing. Day 7 and 10 adult animals showed increased fractional quiescence compared to day 1 adults, while days 2, 3, and 4 adults were generally indistinguishable from day 1 adults (Figure 6(A)). No dramatic differences in bout duration were seen between different aged animals. However, bout number was increased from day 7 at hours 2, 3, and 6 and increased in day 10 adults at hours one, two, three, five, and six (compared to day 1; Figure 6(B,C)). On days 3, 4, 7, and 10, animals began cycling between activity and inactivity more quickly than day 1 adults (Figure 6(D)). A similar decrease in beat rate per minute was also observed at days 2, 3, 4, 7, and 10 (Supplementary Figure 5). These results suggest that locomotory output declines in aged animals, consistent with diminished muscle function in aging animals. Additionally, we noted that different aspects of EIQ metrics decline with age at different rates; time to first EIQ bout decreased with age more rapidly than fractional EIQ increased with age.

Discussion

Here, we work with the computer vision programs poseEIQ and edgeEIQ, which reveal in finer detail changes in *C. elegans* locomotion after prolonged periods of swimming.

Increased inactivity was observed after extended swimming, as were differences in swimming locomotory behaviors between wild-type and mutant animals. Loss of the EGL-4 cGMP-dependent kinase and DAF-16 FOXO function altered EIQ after prolonged swimming. However, loss of the proteins APTF-1 and CEH-17, which are required for developmentally timed sleep and stress-induced sleep in *C. elegans*, respectively, did not affect EIQ. Aged animals showed increased EIQ. Based on examining pharyngeal pumping, animals are actively feeding during the majority of EIQ bouts, indicating that EIQ is usually not a sleep state. Computer-vision programs used here enabled in-depth analysis of EIQ and locomotory behaviors across time and revealed previously undescribed defects. This work and development of these automated analysis strategies enables future work that will interrogate the molecular pathways underlying behaviors associated with exercise and fatigue.

C. elegans locomotory behavior after prolonged swimming has not been thoroughly studied. Previous studies were limited by reliance on manual annotation, which hinders research depth, or by reliance on constrictive microfluidic devices, which may induce mechanical stress (Ghosh & Emmons, 2008; Gonzales, Zhou, Fan, & Robinson, 2019). Using both of our new systems, we can provide a detailed analysis of how *C. elegans* locomotory behaviors change after prolonged swimming. When comparing wild type and *egl-4* mutant animals, differences were found in multiple

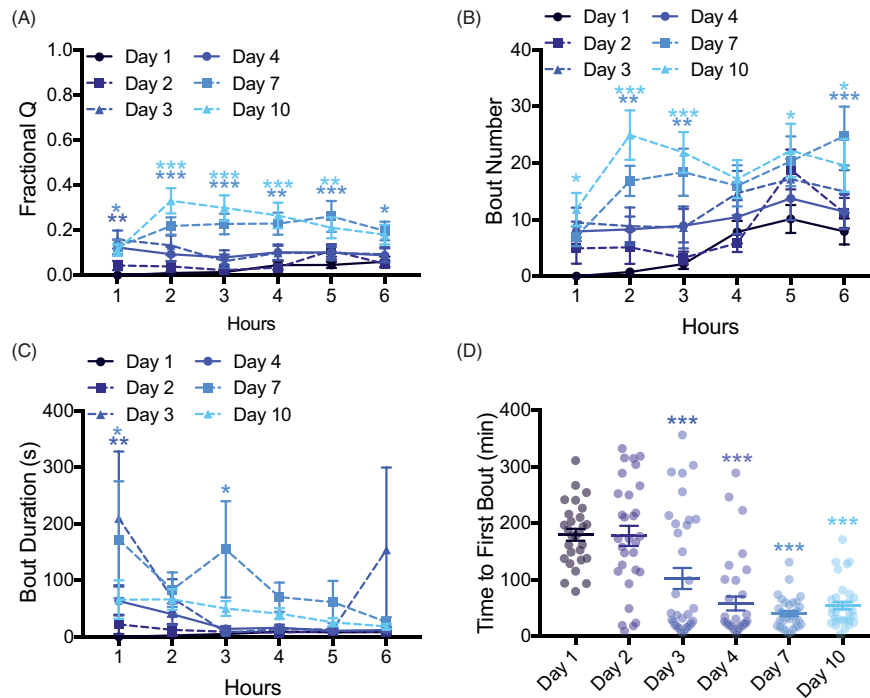


Figure 6. Analysis of exercise-induced quiescence in aged animals. Wild-type animals were aged 1, 2, 3, 4, 7, and 10 days into adulthood. (A) Day 3 adult animals showed increased average fractional quiescence, compared to day 1 adult animals at hour 1. Compared to day 1 adult animals, day 7 adult animals showed increased average fractional quiescence at all time points, while day 10 animals showed increased average fractional quiescence at hours 2 through 5. (B) An increase in average number of bouts per hour, compared to day 1 adult animals, was observed at day 7 adult animals during hours 2, 3, and 6. Day 10 adult animals also showed increased average number of bouts (per hour) during hours 1, 2, 3, 5, and 6, compared to day 1 adult animals. (C) No difference in average bout duration was found amongst the different age groups, with the exception of increased average bout duration in day 3 adult animals at hour 1 and at day 7 adult animals at hours 1 and 3. Two-way ANOVA and Dunnett's multiple comparisons test. (D) Day 3, 4, 7, and 10 adult animals all showed decreased average time to first bout, compared to day 1 adult animals. Kruskal–Wallis test and Dunn's multiple comparisons test. $n = 36$ per group. Error bars indicate \pm SEM. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

parameters, including wave initiation rate at early and late time points. Interestingly, there were also differences in which locomotion parameters changed over time between wild type and mutant animals. For example, curling activity and stretch were found to change over time in wild type animals, but not in *egl-4* mutant animals. In future studies of *C. elegans* fatigue, parameters like wave initiation rate and brush stroke will likely provide information about how vigorously an animal swims and can be used to track decreased muscle output after prolonged swimming exercise. We note that quiescence levels can vary across experiments, even in wild type animals. This may be caused by differences in ambient conditions during rearing (e.g. humidity levels, vibration). Here, all experimental groups in a trial were reared in tandem to control for these differences.

It is important to note that poseEIQ, used herein for analysis of swimming locomotory behaviour, is computationally expensive and requires substantial time for processing. To increase efficiency, we developed edgeEIQ which is faster, but less comprehensive, as it only distinguishes active and inactive states during swimming. Loss of function of the gene *egl-4* has previously been associated with decreased quiescence after prolonged swimming (Ghosh & Emmons, 2008, 2010). To test whether edgeEIQ could detect the impact of *egl-4* mutations on EIQ, we analyzed the behavior

of gain and loss of function mutants of *egl-4*. As expected, gain and loss of function mutations in *egl-4* were associated with increased and decreased EIQ activity, respectively. cGMP-dependent protein kinase EGL-4 is also known to promote quiescent activity during all known forms of *C. elegans* sleep (Hill *et al.*, 2014; Raizen *et al.*, 2008). We originally hypothesized that the quiescent behavior after prolonged swimming might also be a sleep state. Usually sleeping animals will stop both feeding and locomotion. But, we found that pharyngeal pumping was usually observed in animals during EIQ bouts, suggesting that EIQ bouts are usually not sleep. We also used genetic strategies to explore the relationship between sleep and EIQ bouts. The RIS neuron is critical for developmentally timed sleep, and function of the APTF-1 transcription factor is required for RIS-mediated sleep induction (Turek *et al.*, 2013). Likewise, the ALA sensory neuron is required for quiescent behavior during stress-induced sleep (Hill *et al.*, 2014). CEH-17 loss alters gene expression in the ALA neuron, and loss of this protein leads to shortened ALA axons and inability to enter a sleep state following cellular stress (Hill *et al.*, 2014; Pujol, Torregrossa, Ewbank, & Brunet, 2000; Van Buskirk & Sternberg, 2010). We tested animals lacking *aptf-1* and *ceh-17* function for defects in EIQ to determine whether exercise-induced locomotion quiescence was mediated by pathways involved in

mediating locomotion quiescence in developmentally timed or stress-induced sleep. We found that these mutant animals had normal locomotion quiescence. Therefore, EIQ and sleep are not identical and are likely mediated by overlapping, but distinct molecular and cellular pathways.

Prolonged exercise is stressful. In *C. elegans*, the transcription factor DAF-16/FOXO localizes to the nucleus during cellular stress, where it upregulates genes involved with stress response, including oxidative stress response genes (Henderson & Johnson, 2001; Murphy *et al.*, 2003). As prolonged swimming by *C. elegans* leads to transcriptional changes in oxidative stress response genes (Laranjeiro *et al.*, 2017), we reasoned that DAF-16 might play a role in EIQ and stress response. Loss of function *daf-16* mutant animals initiated EIQ earlier than wild type animals, suggesting that DAF-16 normally promotes expression of genes that allow animals to endure the stress of exercise for a longer initial swim period of time.

As they age, animals experience deterioration that can result in fatigue, weakness, and sarcopenia. We examined EIQ as *C. elegans* age and, as predicted, older animals showed increased EIQ. Surprisingly, a dramatic decrease in initial swim period time was seen in animals three or more days into adulthood. This deterioration is not mirrored in fractional EIQ, as animals only show an increase in fractional EIQ at seven or more days into adulthood. Further analysis of EIQ should provide insight into the effects of the aging process on muscle function over time. Changes in EIQ in aged *C. elegans* are likely a marker of healthspan and the mechanisms underlying these changes may be conserved mechanisms relevant to fatigue in aging humans.

Here, we developed systems for automatic analysis of *C. elegans* swimming locomotory behavior with a long-range goal of understanding locomotion quiescence and associated mechanisms. The cellular and molecular pathways that communicate fatigue from muscles to the nervous system during exercise remain obscure. In complex animals, afferent neuronal pathways are thought to carry information from the periphery to the central nervous system, which eventually results in a 'feeling of exhaustion' that results in decreased locomotion. Energy depletion in peripheral organs could also result in decreased locomotion. Results presented here and other works (Beron *et al.*, 2015; Lesanpezeshki *et al.*, 2019; Rahman *et al.*, 2018) have demonstrated that invertebrates show aspects of fatigue, even though they lack extensive afferent neuronal pathways. Further dissection of these behaviors in invertebrates should provide insight into conserved cellular and molecular pathways involved in fatigue, as well as other aspects of endurance and exercise.

Conclusions

Computer-vision systems have been developed for accurate analysis of *C. elegans* locomotory behavior during prolonged swimming exercise (over 6 h). This system is complemented by a faster edge detection-based system that can detect pauses in locomotion. We find that most prolonged swimming results in exercise-induced quiescence (EIQ) bouts that

are dependent on EGL-4/PKG function, but not on the function of genes that are specifically required for *C. elegans* sleep. The timing of EIQ bout initiation is dependent on DAF-16/FOXO function. As *C. elegans* age, the timing, duration, and frequency of EIQ bouts change, consistent with diminished healthspan.

Acknowledgements

The authors acknowledge the Cloud TPU hardware resources that Google made available via the TensorFlow Research Cloud (TFRC) program. Some strains were provided by the CGC, which is funded by the NIH Office of Research Infrastructure Programs (P40 OD010440).

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This research was supported by funds from Brown's Office for the Vice-President for Research (A.C.H. and T.S.). Additional support provided by the Carney Institute for Brain Science, the Center for Vision Research (CVR) and the Center for Computation and Visualization (CCV) at Brown University and a Karen T. Romer Undergraduate Teaching and Research Awards (S.K.).

ORCID

Anne C. Hart  <http://orcid.org/0000-0001-7239-4350>

References

- Beron, C., Vidal-Gadea, A.G., Cohn, J., Parikh, A., Hwang, G., & Pierce-Shimomura, J.T. (2015). The burrowing behavior of the nematode *Caenorhabditis elegans*: a new assay for the study of neuromuscular disorders. *Genes, Brain, and Behavior*, 14(4), 357–368. doi:10.1111/gbb.12217
- Brenner, S. (1974). The genetics of *Caenorhabditis elegans*. *Genetics*, 77(1), 71–94.
- Brown, A.E.X., Yemini, E.L., Grundy, L.J., Jucikas, T., & Schafer, W.R. (2013). A dictionary of behavioural motifs reveals clusters of genes affecting *Caenorhabditis elegans* locomotion. *Proceedings of the National Academy of Sciences of the United States of America*, 110(2), 791–796. doi:10.1073/pnas.1211447110
- Driver, R.J., Lamb, A.L., Wyner, A.J., & Raizen, D.M. (2013). DAF-16/FOXO regulates homeostasis of essential sleep-like behavior during larval transitions in *C. elegans*. *Current Biology*, 23(6), 501–506. doi:10.1016/j.cub.2013.02.009
- Ghosh, R., & Emmons, S.W. (2008). Episodic swimming behavior in the nematode *C. elegans*. *The Journal of Experimental Biology*, 211(Pt 23), 3703–3711. doi:10.1242/jeb.023606
- Ghosh, R., & Emmons, S.W. (2010). Calcineurin and protein kinase G regulate *C. elegans* behavioral quiescence during locomotion in liquid. *BMC Genetics*, 11, 7. doi:10.1186/1471-2156-11-7
- Gomez-Marin, A., Partoune, N., Stephens, G.J., & Louis, M. (2012). Automated tracking of animal posture and movement during exploration and sensory orientation behaviors. *PLoS One*, 7(8), e41642. doi:10.1371/journal.pone.0041642
- Gonzales, D.L., Zhou, J., Fan, B., & Robinson, J.T. (2019). A microfluidic-induced *C. elegans* sleep state. *Nature Communications*, 10(1), 5035. doi:10.1038/s41467-019-13008-5

- Guo, Y., Govindarajan, L., Kimia, B., & Serre, T. (2018). Robust pose tracking with a joint model of appearance and shape. arXiv, 1-11. arXiv:1806.1.
- Henderson, S.T., & Johnson, T.E. (2001). *daf-16* integrates developmental and environmental inputs to mediate aging in the nematode *Caenorhabditis elegans*. *Current Biology*, 11(24), 1975-1980. doi:10.1016/S0960-9822(01)00594-2
- Herndon, L.A., Schmeissner, P.J., Dudaronek, J.M., Brown, P.A., Listner, K.M., Sakano, Y., ... Driscoll, M. (2002). Stochastic and genetic factors influence tissue-specific decline in ageing *C. elegans*. *Nature*, 419(6909), 808-814. doi:10.1038/nature01135
- Hill, A.J., Mansfield, R., Lopez, J.M.N.G., Raizen, D.M., & Van Buskirk, C. (2014). Cellular stress induces a protective sleep-like state in *C. elegans*. *Current Biology*, 24(20), 2399-2405. doi:10.1016/j.cub.2014.08.040
- Huang, H., Singh, K., & Hart, A.C. (2017). Measuring *Caenorhabditis elegans* sleep during the transition to adulthood using a microfluidics-based system. *Bio-protocol*, 7(6), e2174. doi:10.21769/BioProtoc.2174
- Iwanir, S., Tramm, N., Nagy, S., Wright, C., Ish, D., & Biron, D. (2013). The microarchitecture of *C. elegans* behavior during lethargus: homeostatic bout dynamics, a typical body posture, and regulation by a central neuron. *Sleep*, 36(3), 385-395. doi:10.5665/sleep.2456
- Jung, S.-K., Aleman-Meza, B., Riepe, C., & Zhong, W. (2014). QuantWorm: a comprehensive software package for *Caenorhabditis elegans* phenotypic assays. *PLoS One*, 9(1), e84830. doi:10.1371/journal.pone.0084830
- Kimia, B.B., Li, X., Guo, Y., & Tamrakar, A. (2019). Differential geometry in edge detection: accurate estimation of position, orientation and curvature. *IEEE Transactions on Pattern Analysis and Machine Intelligence*, 41(7), 1573-1586. doi:10.1109/TPAMI.2018.2846268
- Kuhn, H.W. (1955). The Hungarian method for the assignment problem. *Naval Research Logistics Quarterly*, 2(1-2), 83-97. doi:10.1002/nav.3800020109
- Laranjeiro, R., Harinath, G., Burke, D., Braeckman, B.P., & Driscoll, M. (2017). Single swim sessions in *C. elegans* induce key features of mammalian exercise. *BMC Biology*, 15(1), 30. doi:10.1186/s12915-017-0368-4
- Lesanpezeshki, L., Hewitt, J.E., Laranjeiro, R., Antebi, A., Driscoll, M., Szewczyk, N.J., ... Vanapalli, S.A. (2019). Pluronic gel-based burrowing assay for rapid assessment of neuromuscular health in *C. elegans*. *Scientific Reports*, 9(1), 15246. doi:10.1038/s41598-019-51608-9
- Marty, E., Liu, Y., Samuel, A., Or, O., & Lane, J. (2017). A review of sarcopenia: enhancing awareness of an increasingly prevalent disease. *Bone*, 105, 276-286. doi:10.1016/j.bone.2017.09.008
- McCloskey, R.J., Fouad, A.D., Churgin, M.A., & Fang-Yen, C. (2017). Food responsiveness regulates episodic behavioral states in *Caenorhabditis elegans*. *Journal of Neurophysiology*, 117(5), 1911-1934. doi:10.1152/jn.00555.2016
- Mulcahy, B., Holden-Dye, L., & O'Connor, V. (2013). Pharmacological assays reveal age-related changes in synaptic transmission at the *Caenorhabditis elegans* neuromuscular junction that are modified by reduced insulin signalling. *The Journal of Experimental Biology*, 216(Pt 3), 492-501. doi:10.1242/jeb.068734
- Murphy, C.T., McCarroll, S.A., Bargmann, C.I., Fraser, A., Kamath, R.S., Ahringer, J., ... Kenyon, C. (2003). Genes that act downstream of DAF-16 to influence the lifespan of *Caenorhabditis elegans*. *Nature*, 424(6946), 277-283. doi:10.1038/nature01789
- Patel, T.P., Gullotti, D.M., Hernandez, P., O'Brien, W.T., Capehart, B.P., Morrison, B., ... Meaney, D.F. (2014). An open-source toolbox for automated phenotyping of mice in behavioral tasks. *Frontiers in Behavioral Neuroscience*, 8, 349. doi:10.3389/fnbeh.2014.00349
- Pujol, N., Torregrossa, P., Ewbank, J.J., & Brunet, J.F. (2000). The homeodomain protein CePHOX2/CEH-17 controls antero-posterior axonal growth in *C. elegans*. *Development*, 127(15), 3361-3371.
- Rahman, M., Hewitt, J.E., Van-Bussell, F., Edwards, H., Blawdziewicz, J., Szewczyk, N.J., ... Vanapalli, S.A. (2018). NemaFlex: a microfluidics-based technology for standardized measurement of muscular strength of *C. elegans*. *Lab on a Chip*, 18(15), 2187-2201. doi:10.1039/C8LC00103K
- Raizen, D.M., Zimmerman, J.E., Maycock, M.H., Ta, U.D., You, Y.-J., Sundaram, M.V., & Pack, A.I. (2008). Lethargus is a *Caenorhabditis elegans* sleep-like state. *Nature*, 451(7178), 569-572. doi:10.1038/nature06535
- Restif, C., Ibáñez-Ventoso, C., Vora, M.M., Guo, S., Metaxas, D., & Driscoll, M. (2014). CeleST: computer vision software for quantitative analysis of *C. elegans* swim behavior reveals novel features of locomotion. *PLoS Computational Biology*, 10(7), e1003702. doi:10.1371/journal.pcbi.1003702
- Schwarz, J., Spies, J.-P., & Bringmann, H. (2012). Reduced muscle contraction and a relaxed posture during sleep-like lethargus. *Worm*, 1(1), 12-14. doi:10.4161/worm.19499
- Stephens, G.J., Johnson-Kerner, B., Bialek, W., & Ryu, W.S. (2008). Dimensionality and dynamics in the behavior of *C. elegans*. *PLoS Computational Biology*, 4(4), e1000028. doi:10.1371/journal.pcbi.1000028
- Swierczek, N.A., Giles, A.C., Rankin, C.H., & Kerr, R.A. (2011). High-throughput behavioural analysis in *C. elegans*. *Nature Methods*, 8(7), 592-598. doi:10.1038/nmeth.1625
- Tramm, N., Oppenheimer, N., Nagy, S., Efrati, E., & Biron, D. (2014). Why do sleeping nematodes adopt a hockey-stick-like posture? *PLoS One*, 9(7), e101162. doi:10.1371/journal.pone.0101162
- Turek, M., Lewandrowski, I., & Bringmann, H. (2013). An AP2 transcription factor is required for a sleep-active neuron to induce sleep-like quiescence in *C. elegans*. *Current Biology*, 23(22), 2215-2223. doi:10.1016/j.cub.2013.09.028
- Van Buskirk, C., & Sternberg, P.W. (2010). Paired and LIM class homeodomain proteins coordinate differentiation of the *C. elegans* ALA neuron. *Development*, 137(12), 2065-2074. doi:10.1242/dev.040881
- Wan, J.-J., Qin, Z., Wang, P.-Y., Sun, Y., & Liu, X. (2017). Muscle fatigue: general understanding and treatment. *Experimental and Molecular Medicine*, 49(10), e384.
- Wei, S.-E., Ramakrishna, V., Kanade, T., & Sheikh, Y. (2016). Convolutional pose machines. arXiv, 1-9. arXiv:1602.00134.
- Yang, Y., & Ramanan, D. (2013). Articulated human detection with flexible mixtures of parts. *IEEE Transactions on Pattern Analysis and Machine Intelligence*, 35(12), 2878-2890. doi:10.1109/TPAMI.2012.261
- You, Y.-J., Kim, J., Raizen, D.M., & Avery, L. (2008). Insulin, cGMP, and TGF-beta signals regulate food intake and quiescence in *C. elegans*: a model for satiety. *Cell Metabolism*, 7(3), 249-257. doi:10.1016/j.cmet.2008.01.005